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# Seascape genomics of common dolphins (*Delphinus delphis*) reveals adaptive diversity linked to regional and local oceanography

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## Abstract

**Background:** High levels of standing genomic variation in wide-ranging marine species may enhance prospects for their long-term persistence. Patterns of connectivity and adaptation in such species are often thought to be influenced by spatial factors, environmental heterogeneity, and oceanographic and geomorphological features. Population-level studies that analytically integrate genome-wide data with environmental information (i.e., seascape genomics) have the potential to inform the spatial distribution of adaptive diversity in wide-ranging marine species, such as many marine mammals. We assessed genotype–environment associations (GEAs) in 214 common dolphins (*Delphinus delphis*) along > 3000 km of the southern coast of Australia.

**Results:** We identified 747 candidate adaptive SNPs out of a filtered panel of 17,327 SNPs, and five putatively locally-adapted populations with high levels of standing genomic variation were disclosed along environmentally heterogeneous coasts. Current velocity, sea surface temperature, salinity, and primary productivity were the key environmental variables associated with genomic variation. These environmental variables are in turn related to three main oceanographic phenomena that are likely affecting the dispersal of common dolphins: (1) regional oceanographic circulation, (2) localised and seasonal upwellings, and (3) seasonal on-shelf circulation in protected coastal habitats. Signals of selection at exonic gene regions suggest that adaptive divergence is related to important metabolic traits.

**Conclusion:** To the best of our knowledge, this represents the first seascape genomics study for common dolphins (genus *Delphinus*). Information from the associations between populations and their environment can assist population management in forecasting the adaptive capacity of common dolphins to climate change and other anthropogenic impacts.

**Keywords:** Delphinids, Adaptive resilience, ddRAD-seq, Landscape genomics, Conservation genomics

## Background

Microevolutionary processes influenced by environmental heterogeneity can create adaptive divergence among populations [1–3]. Marine ecosystems are

environmentally heterogeneous, with coastal and pelagic species impacted by contrasting selective pressures that can lead to local adaptation (e.g., [3–5]). Local adaptation occurs when an individual or group of individuals display higher fitness in a distinct spatial and temporal environment due to specific genetic variants [6–8]. Natural selection acts on both new mutations and standing genetic variation, with most adaptations involving multiple loci and genomic regions [9, 10]. In marine environments, understanding the influence of geomorphological

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and oceanographic features, as well as anthropogenic pressures on genomic variation, enhances our ability to refine knowledge about population structure and rapid evolution [11, 12], including for widespread species with high dispersal potential.

Genomic data has improved our understanding of macro- and microevolutionary processes, providing greater power and accuracy to detect large scale molecular adaptations, as well as population structure, gene flow and adaptive divergence between populations [13–15]. In toothed whales (Odontoceti), most studies of adaptations using genomic markers have focused on a macroevolutionary perspective, while studies investigating ecological specialisation on a microevolutionary level have been documented only for a few species. This includes ecotype adaptations of killer whales (*Orcinus orca* [16]), spinner dolphins (*Stenella longirostris* [17]), finless porpoises (*Neophocaena phocaenoides* [18, 19]), and bottlenose dolphins (*Tursiops aduncus* [20], and *T. truncatus* [21]). Despite these examples, population-level studies of microevolutionary processes remain highly under documented in Odontocetes, specifically in small cetaceans. In particular, little is known about the adaptive resilience of small cetaceans to local or regional environmental changes and to future climatic scenarios. The lack of such studies constrains our capacity to provide information for conservation and management, as well as clarifying important aspects of a species' biology.

The common dolphin (*Delphinus delphis*) is a widespread small cetacean that inhabits temperate, subtropical and some tropical waters around the world [22–24]. Their broad distribution suggests that several habitats are suitable for this species (e.g., [25, 26]). In Australia, common dolphins range from embayments and gulf waters to coastal, shelf and pelagic waters [26–28]. From a neutral genomic perspective, the species in Australasia displays a hierarchical metapopulation structure and fine-scale population sub-structuring [29]. Although common dolphins exhibit high potential for dispersal, prey distribution has been suggested as a main driver for their movements [27, 30, 31]. In Australasian waters, they mainly hunt and feed upon schooling fish such as jack mackerel (*Trachurus declivis*, *T. s. murphyi* and *T. novaezelandiae*), blue mackerel (*Scomber australasicus*), sardines (*Sardina sagax*) and anchovies (*Engraulis australis*) [32, 33]. The ranges of common dolphin populations seem to be influenced by the distribution and abundance of their prey, and often coincide with oceanographic circulation, areas of high primary productivity, and regions of high salinity and low sea surface temperature interfaces [27, 31, 34]. This suggests that oceanographic features and oceanic circulation patterns could be shaping dispersal of common dolphins, as described

for other Australian marine taxa (e.g., [35–37]). However, associations between environmental variables and genetic populations of common dolphin have only been described at broad geographical scales between different oceans [34], and the impact of regional oceanographic features is yet to be revealed. Common dolphins in southern Australia are subject to various anthropogenic stressors, such as interactions and mortalities in fisheries (e.g., [28, 38, 39]), and climatic change (e.g., [40, 41]), both of which can lead to negative health outcomes and potential declines of populations (e.g., [42, 43]). The widespread distribution of common dolphins in southern Australia, where marine environmental gradients and discontinuities are observed, provides an excellent opportunity to investigate microevolutionary processes and adaptive divergence in a highly-mobile marine species.

The temperate waters of southern Australia harbour productive habitats for common dolphins (e.g., [31, 38, 44]). Australia's southern zonal coastal boundary stretches for > 3000 km, with high species endemism [45]. The geographic discontinuity along the large extent of the southern coastal and shelf waters is mainly characterised by: (1) geological features such as > 400 canyons, one bight of > 1000 km, two large inverse estuaries, and a shallow strait; and (2) oceanographic features such as complexly variable bathymetry, strong currents, presence of seasonal upwellings, and gradients in current velocity, salinity and temperature (e.g., [46–48]). These features, which vary from west to east, have been reported to influence historical genetic and genomic subdivision of invertebrate and fish species (e.g., *Nerita* spp. and *Siphonaria* spp. [49], *Chlorophyta* spp., *Phaeophyta* spp. and *Rhodophyta* spp. [50], and *Catomerus polymerus* [51]). Oceanographic and geological characteristics also impact different plankton biomasses (e.g., [52]), which small pelagic fish feed upon [46]. In turn, restrictions in the distribution of these primary producers and consumers may indirectly impact the genetic variation and adaptive potential of large marine predators, including common dolphins, which feed upon small pelagic species across the region.

Seascape genetic/genomics assessments combining genotype and environment associations in marine systems, have the potential to clarify the relative influence of environment and space on genomic variation [3, 53, 54]. Previous studies on population genetic structure of common dolphins in southern Australia based on neutral markers (e.g., mtDNA, microsatellites) [28, 31] and putatively neutral SNPs [29] have hypothesised that common dolphin populations are associated with environmental gradients. For other marine species, seascape genomic/genetic analyses have suggested that adaptive population structure may be driven by environmental gradients of

bathymetry, temperature, oxygen, and salinity (e.g., [53, 55]), while for common dolphins, associations with temperature and chlorophyll have been proposed at the scale of oceanic basins [34].

In this seascape genomics study, genome-wide and environmental data were used to identify loci under selection, and to assess putatively adaptive population structure and diversity along southern Australia. We hypothesised that the continuous distribution of common dolphins in the highly heterogeneous coast of southern Australia influences the genomic variation of the species across different bioregions, leading to adaptive divergence among populations associated with geological and oceanographic features. Findings here can inform about the number and distribution of common dolphin populations and assist with the conservation and management of the species across the region, where it is subject to fisheries interactions, and other anthropogenic impacts such as pollution and climate change (e.g., [38, 56]).

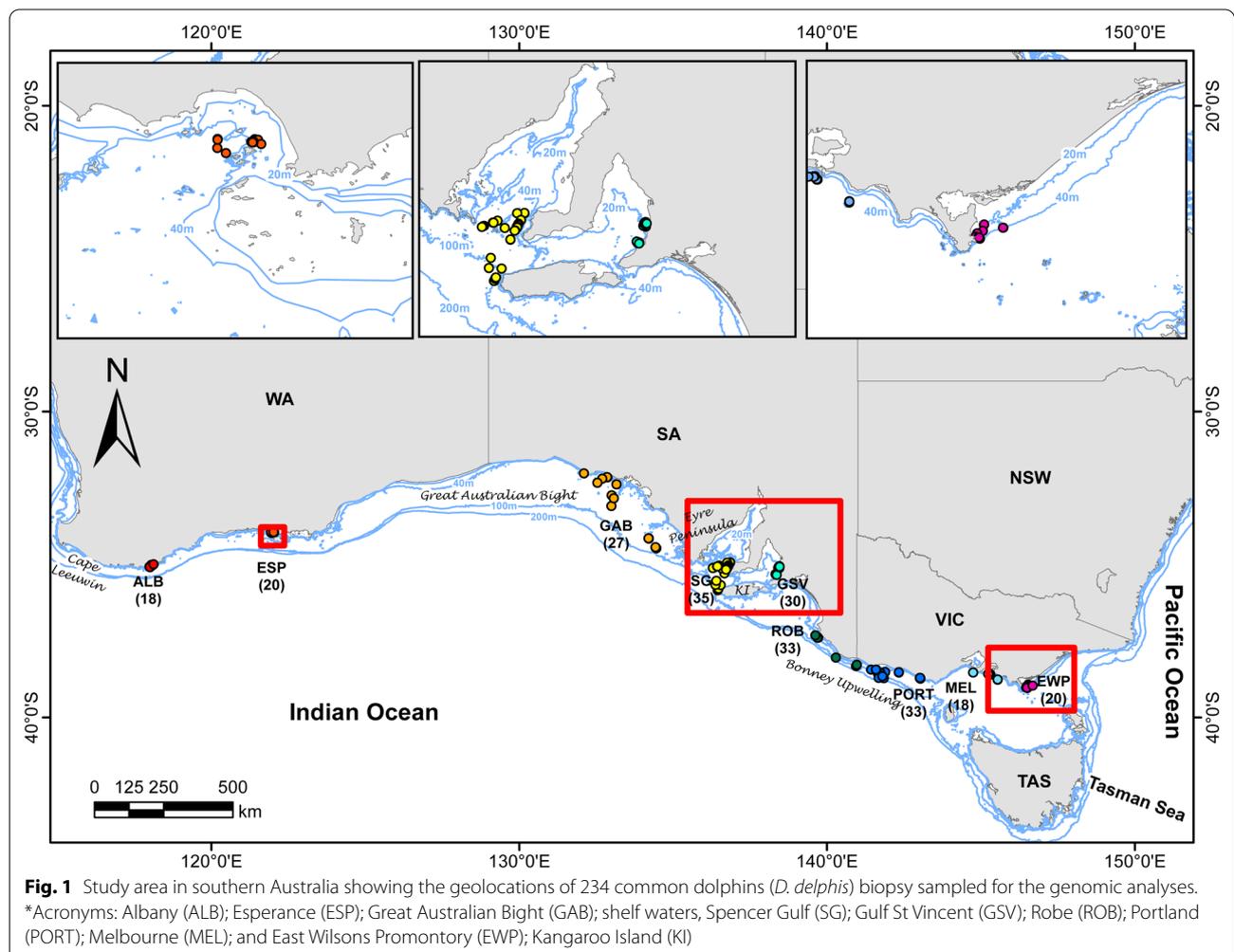
## Results

### Population genomic dataset

A total of 234 biopsy samples of common dolphins were used and sequenced across four Illumina HiSeq 2500 lanes, producing 400 million filtered sequence reads. After filtering using stringent criteria (detailed in Additional file 1: Table S1 and [29]), we obtained a high-resolution dataset of 17,875 filtered SNPs with 1% average missing data per locus. Low-quality individuals, replicates and close relatives ( $|R| \geq 0.5$ ) were removed, resulting in a dataset of 214 individuals for analyses. This dataset was then filtered for Minor Allele Count (MAC) < 3, resulting in a final dataset of 17,327 SNPs (Fig. 1).

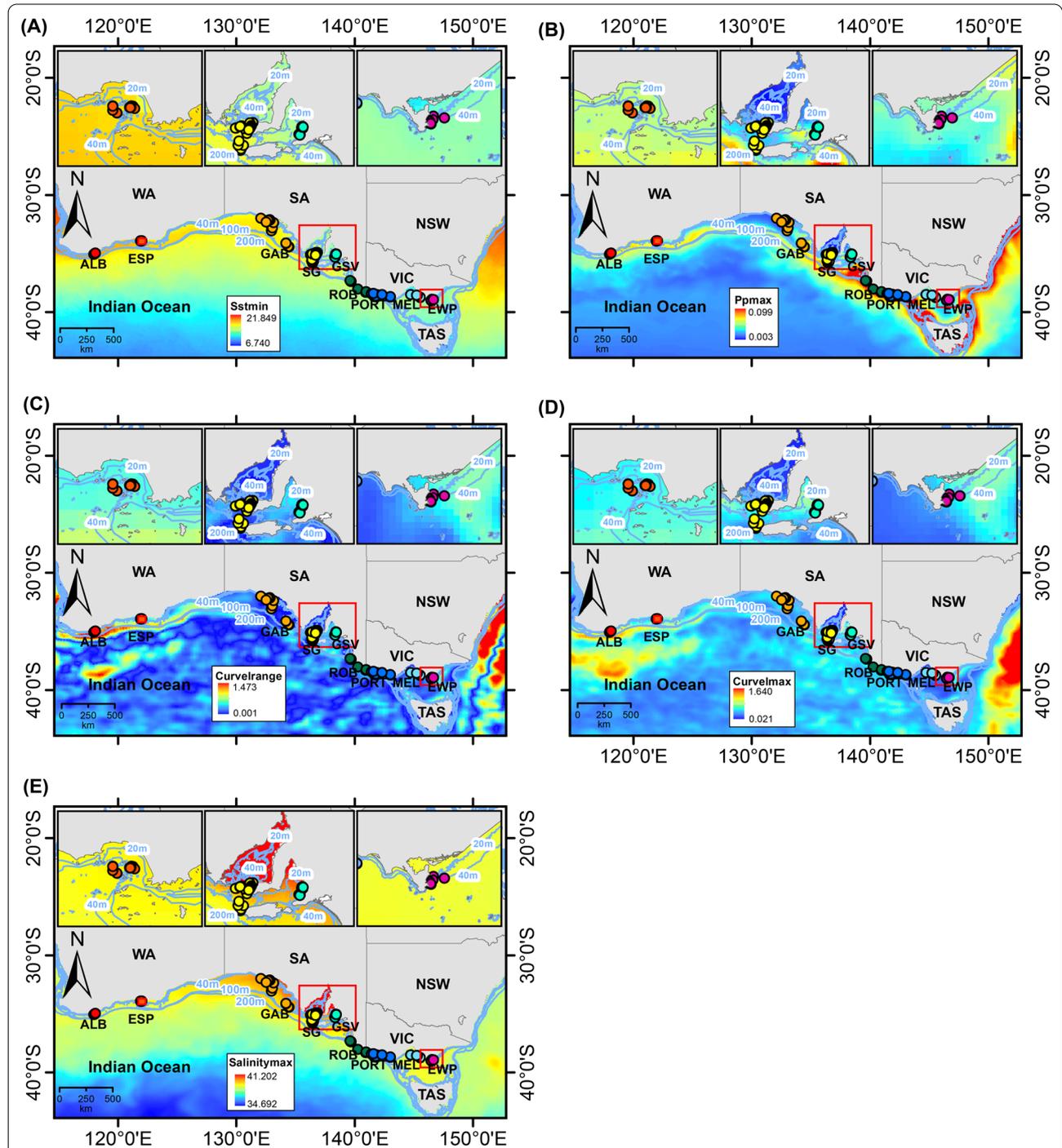
### Genotype-environment associations

A total of six significant MEMs ( $\alpha < 0.01$ ) were retained and used as spatial variables. Out of 24 environmental variables initially included, five were selected after testing pairwise correlation and multicollinearity ( $|r| > 0.7$  and  $VIF \geq 3$ ) (Additional file 1: Fig. S1). The retained



variables were salinity maximum, sea surface temperature minimum, primary productivity maximum, and current velocity maximum and range, with each

environmental variable showing a marked gradient along southern Australia's coast and shelf waters (Fig. 2a–e).

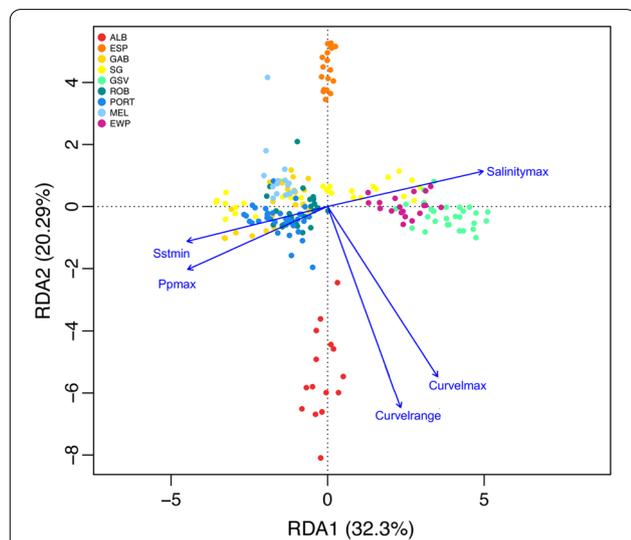


**Fig. 2** Environmental variables that were retained as significant for the Genotype- Environment and Redundancy Canonical Analyses for southern Australian common dolphins (*D. delphis*). **A** Sea surface temperature minimum, **B** Primary productivity maximum, **C** Current velocity range, **D** Current velocity maximum, and **E** Salinity maximum. \*Acronyms as in Fig. 1

The overall RDA model was significant ( $p=0.001$ ), with the spatial variables explaining 4.9% of the variation of the full model, while the genotype dataset and the environmental variables explained 3.5% of the variation. Each of the five retained environmental variables were significant ( $p=0.001$ ) (Additional file 1: Table S3). A total of 747 SNPs were retained as candidate adaptive markers. The first component explained 32% of the constrained variance, while the second component explained 20% of the constrained variance. The RDA biplot demonstrates variation in the genomic response to the five environmental variables among sampling locations (Fig. 3). Changes in primary productivity, sea surface temperature and salinity explained most of the genomic divergence of common dolphins from the Great Australian Bight (GAB), Spencer Gulf (SG), Robe (ROB), Portland (PORT), Melbourne (MEL), East Wilsons Promontory (EWP) and Gulf St Vincent (GSV), with the latter two impacted primarily by higher salinity. In contrast, current velocity variables were strongly associated with the genomic differentiation of common dolphins between the two geographically close sites of Albany (ALB) and Esperance (ESP), Western Australia.

**Adaptive population genomic structure and diversity**

The inferred levels of putatively adaptive genome-wide diversity were relatively high for all sites ( $H_E$  0.369–0.405;  $H_O$  0.361–0.402) compared to the neutral



**Fig. 3** Redundancy Canonical Analysis (RDA) displaying the influence of five environmental variables on individual genomic variation of common dolphins (*D. delphis*) from southern Australia. Legend displays sampling sites from west to east, and colours correspond to where common dolphins were sampled. \*Acronyms as in Fig. 1

**Table 1** Measures of genomic diversity by sampling site based on 747 putatively adaptive SNPs (this study), and 14,799 putatively neutral SNPs (Barceló et al., 2021) from southern Australian common dolphins (*Delphinus delphis*)

Site	N	Neutral		Adaptive	
		$H_O$	$H_E$	$H_O$	$H_E$
ALB	15	0.167	0.166	0.399	0.400
ESP	18	0.171	0.172	0.401	0.403
GAB	22	0.170	0.176	0.399	0.416
SG	32	0.172	0.172	0.419	0.419
GSV	28	0.154	0.160	0.380	0.387
ROB	31	0.172	0.175	0.404	0.413
PORT	32	0.169	0.175	0.406	0.417
MEL	16	0.169	0.177	0.395	0.413
EWP	20	0.175	0.181	0.388	0.406
Total average		<i>0.169</i>	<i>0.173</i>	<i>0.399</i>	<i>0.408</i>
Total SD		<i>0.006</i>	<i>0.006</i>	<i>0.011</i>	<i>0.010</i>

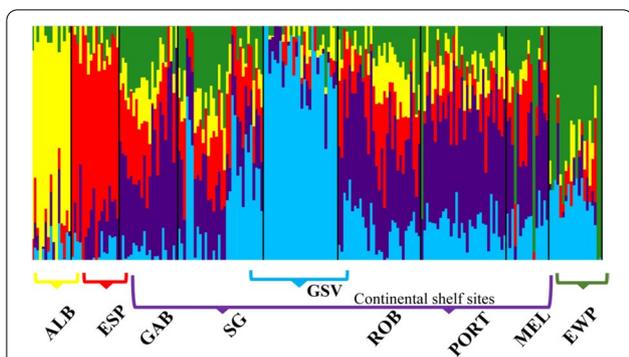
Observed heterozygosity ( $H_O$ ), expected heterozygosity ( $H_E$ ) and number of samples used after filtering adaptive dataset (N). \*Acronyms: Albany (ALB); Esperance (ESP); Great Australian Bight (GAB); shelf waters, Spencer Gulf (SG); Gulf St Vincent (GSV); Robe (ROB); Portland (PORT); Melbourne (MEL); and East Wilsons Promontory (EWP)

Italics show the overall total and standard deviation across sampling sites

genomic diversity ( $H_E$  0.160–0.181;  $H_O$  0.154–0.175) previously reported [29] (Table 1).

Multiple analyses (described below) using the adaptive dataset indicated the presence of four to five local putative populations, which were supported by the AIC test (Additional file 1: Fig. S2): (1) Albany (ALB), (2) Esperance (ESP), (3) Continental shelf sites (GAB, SG, ROB, PORT and MEL), (4) Gulf St Vincent (GSV), and (5) East Wilsons Promontory (EWP). Specifically, Admixture analysis revealed up to five putatively adaptive populations, with a separation between ALB and ESP, GSV, and EWP, compared to considerable admixture among the other sites (Fig. 4; Additional file 1: Fig. S3a–f). PCA results mostly supported four populations, showing in the first two axes only a subtle separation between EWP, GSV sites and [ALB with ESP], with admixed individuals from ALB, ESP and the continental shelf sites clustering in the middle of the two axes, while the third axis show support for a subtle separation between sites of ALB and ESP, with the latest showing a closer association with continental shelf sites (Additional file 1: Fig. S4a–c).

Fixation indices indicated low to moderate ( $F_{ST}$  0.001 to 0.117) genomic differentiation between sampling locations based on the putatively adaptive dataset, with the majority significant and higher than observed with the putatively neutral dataset [29] (see Additional file 1: Fig. S5; Table S4). The greater  $F_{ST}$  differentiation was



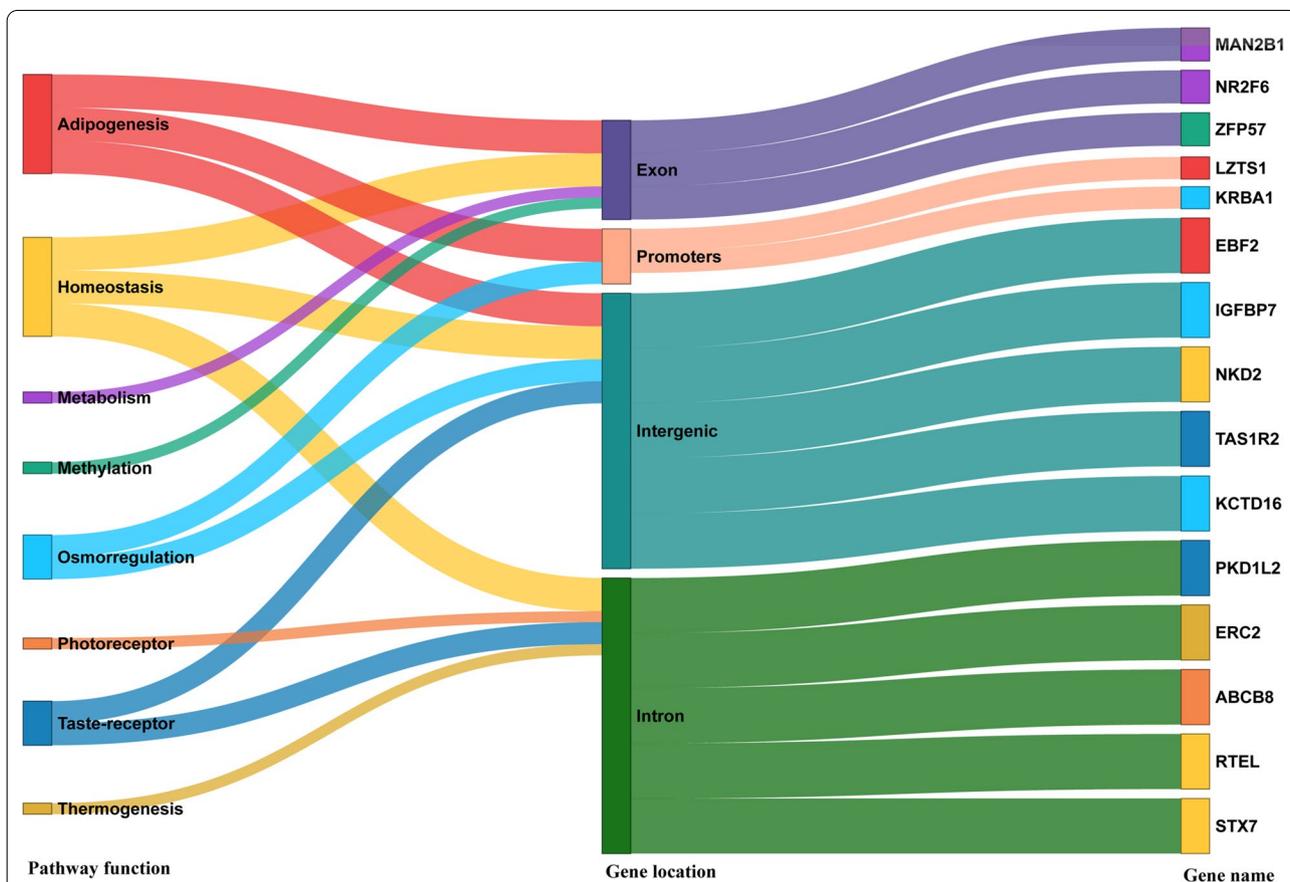
**Fig. 4** Population genomic structure analysis using Admixture based on 747 putatively adaptive SNPs for southern Australian common dolphins (*D. delphis*), labelled by sampling site. The results depict levels of admixture for each individual sample, grouping them into five adaptive genomic clusters (K=5). Each sample is represented by one vertical line and is colour-coded based on the membership probability to one of the identified locally adapted populations. \*Acronyms as in Fig. 1

between GSV and EWP compared to the western sites [ALB and ESP], with the lowest  $F_{ST}$  values showcasing a gradient along southern Australian sites on the continental shelf.

**Functional enrichment and annotation**

A total of 1871 SNPs, from the full dataset of 17,327 SNPs (>10%), scored BLAST hits to the publicly available cetacean nucleotide and non-redundant protein databases (NCBI). Of the 747 potentially adaptive SNPs, 148 were annotated (~19%). Functional enrichment analysis identified 22 GO terms over-represented in the putative adaptive loci (Additional file 1: Table S5). These over-represented GO terms belong mostly to biological processes, with 26 adaptive candidate SNPs in 15 known genes and associated to multiple pathways (Fig. 5).

These 26 SNPs were further investigated, with three of them occurring in exonic regions (Figs. 5; Additional file 1: Fig. S6; Table S6). These SNPs corresponded to candidate genes MAN2B1, which is related to breaking complex sugar molecules in carbohydrate metabolism



**Fig. 5** Sankey diagram showing the 15 of the 26 candidate genes disclosed by the functional enrichment analysis, their interlinkage pathways, and gene location from this study. Colours of each gene corresponds to the function pathway and colours of gene location are only displayed for visual representation

[57–59], and ZFP57 which is related to early embryonic methylation that is potentially altered by nutrients in one's diet [60–62]. These SNPs were most strongly associated with primary productivity variation in the RDA. The third SNP in an exonic region correspond to candidate gene NR2F6 (also known as CoupTFII), which is related to the regulation of adipogenesis, glucose, homeostasis and energy metabolism [63–65], and was correlated with the salinity gradient in the RDA. Although the correlations between candidate SNPs and the environmental variables were generally low ( $r^2 < 0.5$ ), the correlations were still significant (Fisher's  $p$ -value  $< 0.05$ ), and considerable variation in allele frequencies of these candidate SNPs were observed across the seascape (Additional file 1: Fig. S7). Other candidate SNPs were located in promoters (LZTS1, KRBA1), intronic regions (PKD1L2, P3H2, SYT6, ERC2, LEF1, ABCB8, RTEL1, TLN2, MTCL1, STX7 and CFAP54), and intergenic parts of putative genes (LZTS1, EBF2, ELL, IGFBP7, KRBA1, NKD2, TAS1R2, MTCL1 and KCTD16).

## Discussion

Species and populations exhibiting high genomic variation have enhanced prospects for long-term persistence [66–68]. However, highly diverse populations inhabiting multiple environments may be subject to disparate selective pressures, which in turn can result in adaptation to particular habitats [2, 3, 69]. Marine predators are expected to be highly impacted by complex and indirect ecological interactions between their prey and habitats (e.g., [70]). For high dispersal marine species, such as common dolphins, little is known about the spatial distribution of adaptive diversity and its association with spatial connectivity and population subdivision. Here, a seascape genomics approach was used to assess the influence of environmental heterogeneity in shaping putatively adaptive divergence in common dolphins from southern Australia. Our analyses identified over 700 SNPs putatively under selection that delineated up to five populations across the region. The seascape genomics approach revealed four key environmental variables (sea surface temperature, primary productivity, current velocity, and salinity) influencing patterns of genomic variation. This genomic signal appears associated with three different oceanographic phenomena in southern Australian coastal and shelf waters: (1) oceanographic circulation patterns in the western region and related differences in current velocity; (2) upwelling hotspots across the central shelf region influencing fluctuations of primary productivity and sea surface temperatures; and (3) protected coastal environments in the central and

eastern bioregions characterised by marked variations in salinity and seasonal circulation patterns.

## Candidate adaptive genomic variation in southern Australian common dolphins

Genomic variation within populations is impacted by demographic history, but also through ongoing selective pressures that will promote or restrict the dispersal of individuals, which may in turn be reinforced by social structure and behaviour [71, 72]. The comparison between candidate adaptive markers (this study) and previously reported neutral markers [29], showed that estimates of genomic variation and differentiation were higher for the adaptive markers. More complex population structure was also revealed, with five supported populations suggested here using the candidate adaptive markers, compared with just two supported for the neutral dataset [29]. While neutral and adaptive genetic signals both provide useful information about population structure, they arise through different evolutionary forces [68, 73], and can inform different aspects of conservation and management requirements [74–76], as discussed below.

Understanding the pressures that impact marine populations is critical for guiding their management [55]. Common dolphins are a widespread species thought to have colonised different coasts and pelagic habitats off Australia during the Pleistocene [77]. During that period, fluctuations in primary productivity may have opened new niches and promoted colonisation of the region by this species [78–80]. However, contemporary environmental pressures may also influence local adaptation of marine populations [66, 81]. This is relevant for common dolphins in southern Australia, which experience ongoing impacts from anthropogenic activities, particularly by-catch in multiple fisheries (e.g., [38, 82, 83]). Common dolphins exhibit two patterns relevant for their management and conservation. From a neutral perspective, based on signals from genomic regions mostly shaped by genetic drift and migration, genetic connectivity is observed over relatively long geographical distances [29]. From an adaptive perspective, signatures of putative selection supporting local adaptation to some embayment environments, which corroborates field observations of site fidelity and year-round residency in these areas (e.g., [44, 84, 85]). Neutral and adaptive loci can disclose a different number of populations for management and conservation [76, 86]. For common dolphins in southern Australia, adaptive loci supported the hypothesis of putative adaptation to local environments or bioregions that may provide long-term evolutionary potential despite the occurrence of gene flow at the metapopulation level, as previously found with neutral loci [29]. Each

local habitat, or bioregion, exhibits different environmental gradients, such as temperature, current velocity, and salinity, which could be impacting common dolphins and their prey distribution in southern Australia. Differentiation between locally adapted populations despite metapopulation gene flow, has been previously described for other marine taxa along southern Australia (e.g., *Haliotis* [36, 37], and *Nerita* [87]), and in another delphinid (e.g., *T. aduncus* [20]). Effects of climate change may differ between adapted populations, with local populations exhibiting low adaptive diversity, such as those in coastal protected environments or embayments, perhaps more vulnerable to such effects (e.g., [20, 40, 88]). Thus, neutral and adaptive variation are both relevant to conservation for maintaining high standing genetic variation and evolutionary potential across the southern Australia's common dolphin metapopulation.

#### Environmental drivers of adaptive differentiation in southern Australian common dolphins

Adaptive differentiation can be driven by various selective pressures. In marine systems, oceanographic features, such as bathymetry, currents, primary productivity, salinity, and temperature may exert selective pressures [4, 55, 89]. Environmental gradients and discontinuities could be creating soft barriers, and in turn, lead to adaptive divergence among marine populations or bioregions (e.g., [4, 36, 90]). Population differentiation in southern Australian common dolphins has been generally associated with their social structure, areas of high primary productivity, as well as abundance and movements of their prey (e.g., [31, 44, 85]), which mainly exhibit passive dispersal during their larvae stage (e.g., [91–93]). Although using a small representation of the genome in the current study led to a significant, albeit small, association between genomic variation with key environmental variables, it does not necessarily fully elucidate the complex scenarios that occur in the marine system. The assessment of common dolphin populations in southern Australia, thus requires a general ecological understanding of this heterogeneous marine system.

At a global scale, a seascape genetics study of common dolphins based on neutral microsatellite markers found evidence for chlorophyll *a* and sea surface temperature variation linked to major population boundaries [34]. Robust analyses in our study based on a large SNP dataset disclosed that common dolphin genomic variation was associated not just with sea surface temperature, but also current velocity, primary productivity (which is generally positively correlated with chlorophyll *a*), and salinity. Our findings suggest that different environmental variables may be acting on common dolphins' genomic variation in different bioregions, playing major roles in

the differentiation of their populations. The ocean circulation in the region, which influences seasonal differences in primary productivity and sea surface temperature, likely results in different levels of plankton biomass (e.g., [52]), and in turn mediates the abundance and distribution of the dolphins' prey.

#### Ocean circulation off southern Western Australia

The western and southern coasts of Western Australia are characterised by the Leeuwin current, which originates in the warm Indo-Pacific Ocean, moving south along the western coast of Australia, and into the southern coast off Cape Leeuwin [94–96]. The warm waters that enter southern Australia, flowing from west to east, create distinct patterns of temperature, primary productivity, and current velocity along the continental shelf [97, 98]. In southern Western Australia, common dolphins from the two geographically close sites of Albany and Esperance (~300 km apart) were found to be differentiated based on the adaptive SNP dataset, and this distinction seems to be mainly driven by oceanographic currents, particularly changes in current velocity in the region (Figs. 2c, d; 3). This result is consistent with a previous microsatellite study that suggested differentiation between dolphins of these two sites [31], although this separation was not disclosed by neutral SNP study [29]. A recent GEA study in a closely related species, the Indo-Pacific bottlenose dolphin (*T. aduncus*), did not disclose similar separation between localities in the southern coast of Western Australia [20]. This is potentially due to difference in habitat use between the two species, with bottlenose being generally found closer to shore as compared to common dolphins that have a more offshore distribution.

In the western region of southern Australia, the Leeuwin current velocity rapidly declines due to geomorphological formations at the ocean floor, such as the presence of canyons and the Recherche archipelago [93, 96, 99]. Models of larval dispersal in some fish species have shown that individuals move along the Leeuwin current, but separate fish aggregations form between Albany and Esperance [100]. Circulation and geomorphological differences characterise each site; Albany with strong mixing of waters outside the embayment, whereas off Esperance waters are more protected because of the archipelago's presence [101, 102]. Common dolphins are known to optimise their energy requirements by having different hunting strategies and preferences for targeting diverse prey species between different sites and seasons [30, 103, 104]. Thus, common dolphins that inhabit Albany and Esperance could be targeting different fish aggregations, which may lead to differential habitat use.

Interpretation of functional implications between SNPs and their environment needs to be made with caution,

especially in marine systems as it is difficult to elucidate all possible oceanographic and demographic scenarios [105, 106]. However, adaptive genomic variation of common dolphins between Albany and eastern sites was evident in SNPs of the candidate genes EBF2, LEF1 and KCTD16. Variation at these genes correlated with current velocity and primary productivity. These genes are known to be linked to pathways of adipogenesis, homeostasis, thermogenesis and osmoregulation, which promote digestion, absorption of carbohydrates, hypoxic conditions, energy conversion, as well as differentiation of brown adipocytes (e.g., [107–109]).

### ***Southern Australia's continental shelf and its upwellings***

The Leeuwin current continues into southern Australia as the Great Australian Bight current, which is characterised by slower flow in an eastward direction following the break of the continental shelf [95, 97, 110]. The sea floor formation of the southern continental shelf of Australia creates a basin known as the Great Australian Bight, which extends several nautical miles from the coast into the continental shelf break [111–113]. While the warm currents tend to follow the continental shelf, there is also a counter-current of cold water, known as the Flinders current, which typically remains off the continental shelf break [97]. During the austral summer, anticyclonic weather favours the replacement of the warm currents by the cold and productive Flinders current, forming coastal upwellings over the continental shelf [46, 47, 95]. The GEA analysis highlighted the influence of these upwellings as demonstrated by the importance of maximum primary productivity and minimum sea surface temperature in shaping the adaptive genomic differentiation of common dolphins. This was particularly reflected in samples from regions of the Great Australian Bight, mouth of Spencer Gulf, Robe, Portland, and Melbourne, which clustered together, a pattern also disclosed in the PCA and Admixture analyses.

Differences in primary productivity and sea surface temperatures have been considered the main forces that drive seasonal upwellings in southern Australia [46, 100, 112]. There are two types of upwelling centres along southern Australia. Large upwellings are represented by the Bonney upwelling located between Robe, South Australia, and Portland, Victoria, and the Tasmanian upwelling located off western Tasmania. There are also smaller upwelling centres that do not follow the classical Eckman model formation, such as the Eyre Peninsula and the Kangaroo Island upwellings [99, 110, 112]. While the smaller upwellings could mainly have an impact upon the spawning of fish species, such as sardines and anchovies [91], the larger upwelling centres attract a high density

of predators, such as dolphins, whales, seals and sharks feeding upon large biomasses of krill and pelagic fish (e.g., [114–116]).

Common dolphins in open and unprotected continental shelf waters (i.e. Great Australian Bight, mouth of Spencer Gulf, Robe, Portland and Melbourne) presented similar Minor Allele Frequency (MAFs) in the SNPs of candidate genes ERC2 (intronic), and MAN2B1 and ZFP57 (exonic) (SNPs correlated to either primary productivity or temperature), compared to other localities (Additional file 1: Figs. S6 and S7). In other taxa, these genes have been associated with heat stress metabolism, the breaking of complex sugar molecules, and the regulation of fatty acids [59, 117, 118].

### ***Protected coastal habitats***

The geomorphology of southern Australia includes several embayment and protected areas [94, 96]. Some of these protected embayments, such as Gulf St Vincent in South Australia, and Port Phillip Bay in Victoria, have been previously described as important year-round habitats for common dolphins [44, 84, 103]. A previous study using a neutral genomic dataset suggested some differentiation between dolphins from Gulf of St Vincent and other areas [29]. In our study, this differentiation was further informed by the GEA analysis, which disclosed that the genomic variation of common dolphins in sheltered waters of Gulf St Vincent and East Wilsons Promontory is apparently driven by variations in salinity, primary productivity and sea surface temperatures. The Gulf St Vincent is a hyper-saline inverse estuary (i.e., salinity increases with distance to the mouth, and evaporation exceeds inflow circulation), with seasonal circulations that create differences in primary productivity and temperatures [96, 119, 120]. Similarly, Wilsons Promontory is a protected area described as a unique biogeographic region between the southern and eastern Australian currents, with seasonal circulation associated with differences in temperature that promotes the formation of seasonal fish assemblages [49, 121, 122].

In southern Australia, common dolphins mainly feed upon sardines (*S. sagax*), anchovies (*E. australis*), and mackerel (*Trachurus* spp.) [33, 123], which are prey species with high energy density. However, for some presumably resident populations in Port Phillip Bay and Hauraki Gulf, New Zealand, it has been suggested that common dolphins may change their target species based on seasonal availability [30, 103]. Bottlenose dolphins that exhibit strong residency in embayment habitats in southern Australia have also shown genomic differentiation associated with salinity, current velocity and temperature (see Pratt et al. [20] for details), variables than influence temporal fish composition in the region (e.g., [119, 120]),

and could create indirect discontinuities in food availability [20]. For bottlenose dolphins, it was hypothesised, based on genes found to be under potential selection, that some physiological adaptations could be occurring at a population level, especially in embayment habitats [20]. For southern Australian common dolphins from coastal protected habitats, especially those from Gulf of St Vincent, there were significant genomic differences by RDA and enrichment analysis in the MAF of SNPs in genes *STX7* and *IGFBP7* (correlated to salinity), which are genes involved in pathways of osmoregulation and other physiological adaptations (e.g., [124–126]). Macro-evolutionary studies of odontocetes have suggested that adaptative divergence mostly occurred during cycles of high productivity [78–80]. This was observed, for example, in members of the *STX* family genes which are positively selected in the macroevolution of marine mammals (e.g., [16, 127, 128]).

In southern Australian embayments, seasonal changes in salinity and temperature are associated with changes in the composition of fish assemblages (e.g., [122, 129]). Genomic variation in common dolphins inhabiting embayment areas, such as Gulf St Vincent, could relate to mechanisms for coping with high salinities, while allowing them to remain locally resident year-round by alternating feeding upon different prey species. Although the evidence reported here is based on a small representation of the genome (less than 2% of the dolphin genome with a 99% alignment to *T. aduncus*), it is expected that future studies using species-specific whole genomes, will expand and report on many other gene regions and pathways likely to be under selection in these common dolphin populations.

#### Implications for conservation under future climatic scenarios

This study points to environmental variables that may be influencing putatively adaptive populations of common dolphins across southern Australia. With rapid and ongoing climatic change and other anthropogenic pressures impacting on marine species [3, 11, 130], it is essential to understand which environmental factors shape genomic variation to identify locally adapted populations relevant for conservation and management. Models predicting the impact of climate change in marine systems have provided evidence that differences in circulation patterns will likely lead to warmer environments [131–133]. For cetaceans, two possible scenarios have been proposed. One scenario suggests that changes in ocean circulation, wind patterns and currents could enhance upwelling areas, with large predators such as common dolphins likely benefiting from these changes (e.g., [41, 134, 135]). In contrast, another scenario suggests that warmer sea

surface temperatures would alter community dynamics and increase exposure of populations to various pathogens (e.g., [40–42]). In this second scenario, prey abundance and distribution could be greatly impacted by effects on plankton biomasses (e.g., [136, 137]), which could lead to disease outbreaks, prey depletion and population declines for cetaceans and other marine predators.

Potential effects suggested in both scenarios could impact southern Australian common dolphins. The first scenario is perhaps most likely for common dolphin populations inhabiting sites where connectivity persists over thousands of kilometres due to seasonal aggregations in the upwelling areas (e.g., [41, 135]). Southern Australian common dolphins could enhance the movement of nutrients to different habitats and trophic levels (e.g., [138, 139]). This movement of nutrients due to climate change could affect the timing and magnitude of the upwellings, which could deplete some areas and eutrophicate others (e.g., [136]), impacting common dolphins and other cetacean species that feed upon high density prey biomasses [137, 139, 140]. In contrast, the second scenario could potentially be more relevant for common dolphins that live in protected habitats in which extreme climatic events, such as marine heatwaves, could lead to high mortalities of prey species (e.g., [141]) and alteration of spawning times (e.g., [92, 142]). Moreover, changes in temperature and nutrients of waters masses could lead to low abundance and redistribution of prey species (e.g., [143, 144]). For dolphin species that inhabit protected environments, epizootic events often coincide with these types of extreme climatic stressors, leading to negative impacts on population health and reproduction, and occasionally large morbidity and mortality events (e.g., [40, 43, 145]). Moreover, common dolphins in embayment areas such as Gulf St Vincent and Spencer Gulf have been subjected to fisheries interactions for long periods of time [28, 38, 56], and extreme climatic events leading to lower food availability may further compound negative impacts (e.g., [40, 43, 140]). The dynamics of marine ecosystems are extremely complex, and future climatic changes may lead to indirect effects that need to be contemplated in future conservation planning [46, 144].

As a near top predator, common dolphins provide important ecosystem services to the marine environment [138, 139], and if further anthropogenic or climatic impacts were to occur, these could lead to changes in food-webs potentially causing the eutrophication of ecosystems (e.g., [40, 146, 147]). Adaptation to heterogeneous environments in species with high genomic diversity can promote population resilience to climatic changes [72]. When developing policies and management decisions, it is important to incorporate information from both neutral and adaptive markers to ensure the

persistence of high standing genomic variation in marine populations [2, 69, 148]. Currently, these putatively divergent dolphin populations are being managed according to the management stocks of their prey (e.g., [149]), with no specific consideration of common dolphin genetic or genomic differentiation. Results of this study disclosed five putatively adaptive common dolphin populations in southern Australia that need to be considered as priority areas for conservation and management, taking into account the potential cumulative impacts of fisheries (e.g., [38]) and other stressors on each local population, as well as across its Australasian metapopulations [29]. In line with Funk et al.'s [86] framework for the delineation of conservation units, we suggest that common dolphin management units should incorporate results of both neutral [29] and adaptive population structure. Integrating these results can lead to management units that enhance functional corridors, and provide long-term, high standing genetic variation to the populations. We also recommend that future common dolphin studies should implement this framework as already used for other marine taxa (e.g., *Parastichopus* [12], *Microtus* [150], *Carcharhinus* [151]). Management units from areas such as Gulf of St Vincent should be prioritised, given it continues to be impacted by human activities, exhibits the least amount of migration compared to adjacent sites [29], and is an area where common dolphins show putative adaptation to the semi-enclosed embayment.

## Conclusion

Our study suggest that conservation and policy efforts towards common dolphins should preserve diversity as well as connectivity, and take into consideration cumulative impacts on the putatively adaptive populations as a proxy of evolutionary potential. The GEA analysis indicated that common dolphin genomic variation is impacted by four key environmental variables, which in turn are likely related to three oceanographic phenomena that characterise this broad ocean region. Genomic variation in dolphins off the southern coast of Western Australia was associated with current velocity, while genomic differentiation of common dolphins from sites along the continental shelf break were associated with primary productivity and sea surface temperature. The latter may relate to major upwelling centres, which could be promoting areas of seasonal aggregation. In contrast, genomic differentiation of common dolphins from protected coastal habitats and embayments were associated mainly with fluctuations in salinity. These environmental variables present gradients and discontinuities, which may create soft barriers among the putative populations. Thus, it is recommended that neutral and adaptive variation should be considered for the management of these

five putatively locally adaptive common dolphin populations, while allowing long-range gene flow to persist across their previously described Australasian metapopulation. Maintaining connectivity can promote long-term high standing genomic variation, which in turn will enhance population viability under anthropogenic impacts, including unfavourable climatic events. Furthermore, this study represents the first seascape genomic assessment of common dolphins in a dynamic heterogeneous environment. The candidate genes described here may be useful for future comparative studies of common dolphins and potentially other delphinid species that share similar environments.

## Methods

### Sample collection and study area

Common dolphins were sampled across >3000 km of southern Australian waters between 2002 and 2015, with locations allocated based on individual GPS data (Fig. 1). These sites encompass the distribution of this species along the oceanographic, environmental, and geological discontinuities of southern Australia. Biopsy samples were collected from live individuals using a hand held biopsy pole [152] or remote biopsy gun (PAXARMS) [153]. Dependent calves were not sampled to avoid the inclusion of closely related individuals. Biopsy samples were preserved in 90% ethanol or in a 20% salt-saturated solution of dimethyl sulphoxide (DMSO) and stored frozen ( $-80^{\circ}\text{C}$ ) until laboratory analyses took place. A total of 234 biopsy samples were used for the genomic analyses.

### Laboratory analyses and bioinformatics

DNA extractions from biopsy samples were performed using a salting-out protocol [154]. Quantity and quality controls of DNA were determined using a Qubit 2.0 fluorometer (Life 178 Technologies), a Nanodrop spectrophotometer (Thermo Scientific), and gel electrophoresis. Library preparation for the double digest restriction-site associated (ddRAD) was performed following the protocol of Peterson et al. [155], and sequencing was done using an Illumina HiSeq 2500 platform producing single-end, 100 bp reads, at the South Australian Health & Medical Research Institute (SAHMRI).

Raw data quality was assessed, and demultiplexed using `process_radtags`, with STACKS v1.48 [156, 157], reads were trimmed by quality (only one error allowed) using TRIMMOMATIC [158], and then dDocent2.2.19 pipeline [159] was used to call the SNPs. The resulting loci were then filtered using VCFtools (see details in Additional file 1 and [29]). The quality-filtered ddRAD loci were then mapped against the genome of a closely related species, the Indo-Pacific bottlenose dolphin (*T. aduncus*)

from southern Australia [145] using Bowtie2 [160], due to the absence of a high-quality *D. delphis* genome. Genotype errors were considered and filtered by the incorporation of 10 known replicate samples. To exclude potential field sample duplicates or closely related individuals in the dataset, relatedness between pairs of individuals was calculated by the triadic likelihood estimator (TrioML) in Coancestry v1.0.1.9, as this estimator provides the highest correlation with true values [161]. Then, following a simulation protocol of Attard et al. (see [162] for details of parameters), we determined a relatedness threshold ( $|R| > 0.5$ ) to remove one individual per each pair of duplicates or closely related individuals, such as parent-offspring (detailed in Additional file 1: Table S1).

#### Selection of environmental variables and spatial data

Bathymetry, sea surface temperature, chlorophyll *a*, current velocity, primary productivity, and salinity were selected as ecologically relevant environmental variables to test for associations with common dolphin genomic variation based on previous studies (e.g., [29, 31]). For each of the variables selected, the annual maximum, mean, minimum, and range values between the years 2000 and 2014 were used, resulting in a total of 24 variables (Additional file 1: Table S2). The environmental data was downloaded from the database BioOracle at a resolution of ~9.2 km, using the R package 'sdmpredictors' [163, 164] (see Additional file 1 for details).

To control for spatial autocorrelation, pairwise oceanic distances were calculated using the GPS coordinates of each individual and the R function *viamaris* available at <https://github.com/pygmyperch/melfuR>. Pairwise oceanic distances were then transformed to Moran's eigenvector maps (MEM) using the package 'memgene' with a forward selection procedure, 100 permutations, and an alpha value of 0.05 [165]. The selected MEMs were then used as the spatial variables for analyses.

To determine which environmental variables were significantly driving the population genomic differentiation from the initial 24 environmental variables, standardisation was first implemented with the R package 'psych' [166], before excluding highly correlated variables ( $r > 0.7$ ) [167–169] and those with a variance inflation factor (VIF)  $\geq 3$  [170]. We then used a forward selection criteria with the R package 'vegan' [171] to retain environmental variables that explained a significant ( $p < 0.05$ ) portion of the genomic variation [166, 172].

#### Genotype-environment association

Loci putatively under selection were detected using a seascape genomics approach within an individual-based genotype-environment association (GEA) framework. This methodology allows the identification of

associations between genetic and selected environmental variables across individuals, with multivariate analyses carried out in the R package 'vegan' [171] (see description and parameters of analysis below). The Redundancy Canonical Analysis (RDA) was chosen because it usually outperforms other GEA methodologies, such as univariate analyses (e.g., Latent Factor Mixed Models), for detecting genomic markers associated with environmental variables, as it reduces the number of false-positives without compromising the detection of true-positive candidates (e.g., [173, 174]). A partial RDA was used to assess the effect of the selected environmental variables on the genomic diversity while controlling for the spatial pattern using the selected MEMs.

The significance of each environmental variable and axis ( $p < 0.05$ ) was calculated using an Analysis of Variance (ANOVA) with 1000 permutations. Loci that scored greater than three standard deviations ( $\pm 3SD$ ) from the mean locus scores were selected as candidates for each of the significant RDA axes. Selected axes explained a significant ( $p < 0.05$ ) portion of the genomic variation, as previously suggested [172, 173]. Spearman's correlations were then calculated between each of the candidate loci and the retained environmental variables to determine the most important environmental variable shaping allele frequencies of each locus.

#### Adaptive population diversity and structure

Genomic diversity of the candidate SNPs was assessed for each sampled site using GenoDive 2.0b27 [175]. Principal Component Analysis (PCA), which is a model-free approach, was used to investigate population structure using the R package 'Adegenet' [176, 177]. The Akaike Information Criterion (AIC) was then used to determine the best-supported number of clusters in the dataset, using the *snapclust.chooseK* function, also in 'Adegenet' [178].

Population structure was further investigated using a Bayesian clustering approach that infers population stratification based on the estimated individual ancestries in Admixture v1.3.0 [179]. Although the candidate adaptive dataset likely violates Hardy–Weinberg assumptions of equilibrium [86], this analysis was used as a comparison to the results based on the putatively neutral dataset [29]. The maximum likelihood estimates were calculated by using the ancestry portion and the population allele frequency to assign the most likely number of *K* (i.e., populations) in the dataset, testing for *K* 1–9, and for modelling the probability of observed genotypes [180]. This was followed by cross validation with ten replicates for each of the *K* values.

Adaptive genomic differentiation among sites was estimated as pairwise  $F_{ST}$  [181] using GenoDive 2.0b27. Significance levels were assessed using 10,000 permutations, corrected by the B-Y method (FDR < 10%; [182]). Heatmap plots of  $F_{ST}$  were constructed with the R package 'ggplot2' [183].

### Functional enrichment analysis and annotation

A functional enrichment analysis was performed for the full annotated dataset of 17,327 SNPs as a background gene set, using the nucleotide and non-redundant protein NCBI databases for all available cetacean sequences [53, 184–186]. Based on the results of linkage disequilibrium (details in [29]), flanking sequences of 300 bp either side of each SNP were extracted, resulting in 601 bp sequences. Annotation was then performed using the alignment tool (BLAST) in the NCBI nucleotide and non-redundant protein databases, with an e-value threshold of  $1 \times 10^{-3}$ . A Gene Ontology (GO) term enrichment analysis was also performed comparing the candidate loci to the full dataset (17,327 SNPs) using a Fisher's exact test and a FDR cut-off of  $\leq 5\%$  [187]. The resulting GO terms related to a specific SNP in a candidate gene, were then further examined for its sequence position using snpEFF [188]. Each SNP was further assigned to a pathway and function using the Reactome [189] and UniProtKB databases, respectively (The UniProt Consortium, 2018).

### Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12862-022-02038-1>.

**Additional file 1: Table S1.** Filtering steps and number of SNPs retained after each step for common dolphins (*Delphinus delphis*) in southern Australia. FDR: false discovery rate. **Table S2.** Summary of environmental variables included in the genotype-environment association multivariable analyses, datasets were retrieved from BioOracle. **Table S3.** (A) Significance of the RDA and the proportion explained by each component of the full model selection. (B) Significance of the environmental variables selected, with an overall significance of the model and variables at  $p = 0.0001$ . **Table S4.** Pairwise  $F_{ST}$  values between sites based on putatively adaptive (this study) and neutral (1) datasets for southern Australian common dolphins (*Delphinus delphis*). Upper right, neutral  $F_{ST}$  values, and lower left adaptive  $F_{ST}$  values, with their significance of the p-values by the B-Y method corrected represented by \*\*\*0.0001, \*\*0.001. Acronyms for sites as in Fig. 1. **Table S5.** Significance of Gene Ontology (GO) terms for southern Australian common dolphins (*Delphinus delphis*), comparing the full dataset with the putative candidate loci by Fisher's exact test. Biological Process (BP), Molecular Function (MF), Cellular Component (CC). **Table S6.** Function of the candidate genes found in exonic regions, which were over enriched by the Gene Ontology analyses, for the 747 putatively adaptive SNPs discovered by the RDA of southern Australian common dolphins (*Delphinus delphis*). **Figure S1.** Multicollinearity between the five environmental variables used for the RDA. Salinity maximum (BO2\_salinitymax\_ss), primary productivity maximum (BO2\_ppmax\_ss), sea surface temperature minimum (BO\_sstmin), current velocity maximum (BO2\_curvelmax\_ss) and current velocity range (BO2\_curvelrange\_ss). Numbers in the upper right matrix are the correlation values of each comparison; the smaller the number gets, the closest to zero the correlation

between the variables is. Variables were standardised from 0-1. **Figure S2.** Akaike Information Criterion (AIC) used to determine the best-supported number of clusters in the PCA, based on the putative adaptive dataset of southern Australian common dolphins (*Delphinus delphis*). **Figure S3.** Population genomic structure analysis using Admixture based on the putatively adaptive SNPs for southern Australian common dolphins (*Delphinus delphis*) (labelled by sampling site and individual). K represents the number of populations tested (A to F), in which K4\* and K5\* are both correctly assigned as they are the most supported and highly likely number of local populations suggested by the analyses. Acronyms for sites as in Fig. 1. **Figure S4.** Principal Component Analysis (PCA) based on 747 candidate adaptive loci for southern Australian common dolphins (*Delphinus delphis*). (A) Explanatory axes PC1 vs PC2. (B) Explanatory axes PC1 vs PC3. (C) Explanatory axes PC2 vs PC3. Acronyms for sites as in Fig. 1. **Figure S5.** Heatmap of pairwise  $F_{ST}$  values between sites based on the adaptive (this study) and neutral (1) SNP datasets for southern Australian common dolphins (*Delphinus delphis*). Upper right, neutral dataset, and lower left, adaptive dataset. Acronyms for sites as in Fig. 1. **Figure S6.** Allele frequency changes for the candidate gene variants found in exonic regions of common dolphins (*Delphinus delphis*) across sampling sites in southern Australia. (A) NR2F6/NR2F2 was associated with maximum salinity, (B) ZFP57, with primary productivity maximum, and (C) MAN2B1, with primary productivity maximum. **Figure S7.** Minor allele frequencies of 26 SNPs selected by a gene enrichment test, with variants in coding or non-coding regions.

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### Author contributions

AB Conceptualization and design of the study, DNA extractions, ddRAD library preparation, bioinformatics, data analyses, writing of the manuscript and corresponding author. LM Conceptualization and design of the study, guidance on lab methods, data analyses and interpretation, collection of samples and drafting of manuscripts. LB Design of the study, guidance on lab methods, data analyses and interpretation, and drafting of manuscripts. KB Collection of samples and drafting of manuscripts. JSC Guidance on laboratory methods, bioinformatics, data analyses and drafting of final manuscript. CB Guidance with data analyses and drafting of final manuscript. GP Collection of samples and drafting of final manuscript. All authors read and approved the final manuscript.

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### Availability of data and materials

The datasets generated and/or analysed during the current study are available in the FigShare repository <https://figshare.com/s/ff6e85619698e8a90b8f3> (<https://doi.org/10.6084/m9.figshare.19319732>).

### Declarations

#### Ethics approval and consent to participate

All institutional and/or national guidelines and regulations regarding the collection of biopsy samples and protocols were followed. This study was conducted with animal ethics approval of the Flinders University and Macquarie University under research permit (E326), as well as research permits from Department of Environment and Natural Resources, South Australia (E25889); Department of Environment and Conservation, Western Australia (SF008961); Department of Sustainability Environment, Victoria (FF383247); Department

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#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that the research was conducted in the absence of any conflict of interest or competing interests.

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#### References

- Balkenhol N, Cushman SA, Storfer A, Waits L. Landscape genetics concepts, methods and applications 2016.
- Manel S, Holderegger R. Ten years of landscape genetics. *Trends Ecol Evol*. 2013;28(10):614–21.
- Grummer JA, Beheregaray LB, Bernatchez L, Hand BK, Luikart G, Narum SR, et al. Aquatic landscape genomics and environmental effects on genetic variation. *Trends Ecol Evol*. 2019;34(7):641–54.
- Benestan L, Quinn BK, Maaroufi H, Laporte M, Clark FK, Greenwood SJ, et al. Seascape genomics provides evidence for thermal adaptation and current-mediated population structure in American lobster (*Homarus americanus*). *Mol Ecol*. 2016;25(20):5073–92.
- Sandoval-Castillo J, Beheregaray LB. Oceanographic heterogeneity influences an ecological radiation in elasmobranchs. *J Biogeography*. 2020;47(7):1599–611.
- Hoban S, Kelley JL, Lotterhos KE, Antolin MF, Bradburd G, Lowry DB, et al. Finding the genomic basis of local adaptation: pitfalls, practical solutions, and future directions. *Am Nat*. 2016;188(4):379–97.
- Savolainen O, Lascoux M, Merila J. Ecological genomics of local adaptation. *Nat Rev Genet*. 2013;14(11):807–20.
- Lotterhos KE, Whitlock MC. The relative power of genome scans to detect local adaptation depends on sampling design and statistical method. *Mol Ecol*. 2015;24(5):1031–46.
- Pritchard JK, Pickrell JK, Coop G. The genetics of human adaptation: hard sweeps, soft sweeps, and polygenic adaptation. *Curr Biol*. 2010;20(4):R208–15.
- Barrett RD, Schluter D. Adaptation from standing genetic variation. *Trends Ecol Evol*. 2008;23(1):38–44.
- Manel S, Loiseau N, Andreollo M, Fietz K, Goni R, Forcada A, et al. Long-distance benefits of marine reserves: myth or reality? *Trends Ecol Evol*. 2019;34(4):342–54.
- Xuereb A, D'Aloia CC, Andreollo M, Bernatchez L, Fortin MJ. Incorporating putatively neutral and adaptive genomic data into marine conservation planning. *Conserv Biol*. 2020;35:909.
- Kelley JL, Brown AP, Therkildsen NO, Foote AD. The life aquatic: advances in marine vertebrate genomics. *Nat Rev Genet*. 2016;17(9):523–34.
- Therkildsen NO, Hemmer-Hansen J, Als TD, Swain DP, Morgan MJ, Trippel EA, et al. Microevolution in time and space: SNP analysis of historical DNA reveals dynamic signatures of selection in Atlantic cod. *Mol Ecol*. 2013;22(9):2424–40.
- Cammen KM, Andrews KR, Carroll EL, Foote AD, Humble E, Khudiyakov JI, et al. Genomic methods take the plunge: recent advances in high-throughput sequencing of marine mammals. *J Hered*. 2016;107(6):481–95.
- Foote AD, Vijay N, Avila-Arcos MC, Baird RW, Durban JW, Fumagalli M, et al. Genome-culture coevolution promotes rapid divergence of killer whale ecotypes. *Nat Commun*. 2016;7:11693.
- Andrews KR, Epstein B, Leslie MS, Fiedler P, Morin PA, Hoelzel AR. Genomic signatures of divergent selection are associated with social behavior for spinner dolphin ecotypes. *Mol Ecol*. 2021.
- Ruan R, Guo AH, Hao YJ, Zheng JS, Wang D. De novo assembly and characterization of narrow-ridged finless porpoise renal transcriptome and identification of candidate genes involved in osmoregulation. *Int J Mol Sci*. 2015;16(1):2220–38.
- Zhou X, Guang X, Sun D, Xu S, Li M, Seim I, et al. Population genomics of finless porpoises reveal an incipient cetacean species adapted to freshwater. *Nat Commun*. 2018;9(1):1276.
- Pratt EAL, Beheregaray LB, Bilgmann K, Zanardo N, Diaz-Aguirre F, Brauer C, et al. Seascape genomics of coastal bottlenose dolphins along strong gradients of temperature and salinity. *Mol Ecol*. 2022.
- Louis M, Galimberti M, Archer F, Berrow S, Brownlow A, Fallon R, et al. Selection on ancestral genetic variation fuels repeated ecotype formation in bottlenose dolphins. 2021;7(44):eabg1245.
- Möller LM. Sociogenetic structure, kin associations and bonding in delphinids. *Mol Ecol*. 2011;21(3):745–64.
- Natoli A, Cañadas A, Vaquero C, Politi E, Fernandez-Navarro P, Hoelzel AR. Conservation genetics of the short-beaked common dolphin (*Delphinus delphis*) in the Mediterranean Sea and in the eastern North Atlantic Ocean. *Conserv Biol*. 2008;9(6):1479–87.
- Perrin WF. World Cetacea Database [marinespecies.org/cetacea2021](http://marinespecies.org/cetacea2021).
- Becker EA, Carretta JV, Forney KA, Barlow J, Brodie S, Hoopes R, et al. Performance evaluation of cetacean species distribution models developed using generalized additive models and boosted regression trees. *Ecol Evol*. 2020;10:5759.
- Bilgmann K, Parra GJ, Möller LM. Occurrence, distribution and abundance of cetaceans off the western Eyre Peninsula in the Great Australian Bight. *Deep Sea Res Part II Top Stud Oceanogr*. 2018;157–158:134–45.
- Möller LM, Valdez FP, Allen S, Bilgmann K, Corrigan S, Beheregaray LB. Fine-scale genetic structure in short-beaked common dolphins (*Delphinus delphis*) along the East Australian Current. *Mar Biol*. 2011;158(1):113–26.
- Bilgmann K, Möller LM, Harcourt RG, Gales R, Beheregaray LB. Common dolphins subject to fisheries impacts in Southern Australia are genetically differentiated: implications for conservation. *Anim Conserv*. 2008;11(6):518–28.
- Barceló A, Sandoval-Castillo J, Stockin KA, Bilgmann K, Attard CRM, Zanardo N, et al. A matter of scale: population genomic structure and connectivity of fisheries at-risk common dolphins (*Delphinus delphis*) from Australasia. *Front Mar Sci*. 2021;8.
- Peters KJ, Bury SJ, Betty EL, Parra GJ, Tezanos-Pinto G, Stockin KA. Foraging ecology of the common dolphin *Delphinus delphis* revealed by stable isotope analysis. *Mar Ecol Prog Ser*. 2020;652:173–86.
- Bilgmann K, Parra GJ, Zanardo N, Beheregaray LB, Möller LM. Multiple management units of short-beaked common dolphins subject to fisheries bycatch off southern and southeastern Australia. *Mar Ecol Prog Ser*. 2014;500:265–79.
- Meynier L, Stockin KA, Bando MKH, Duignan PJ. Stomach contents of common dolphin (*Delphinus* sp.) from New Zealand waters. *N Z J Mar Freshwater Res*. 2008;42(2):257–68.
- Goldsworthy SD, Ferguson GJ, Ward TM. Assessment of dolphin interactions, effectiveness of Code of Practice and fishing behaviour in the South Australian Sardine Fishery: 2018–19. In: Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences) A, editor. F2010/000726-10. SARDI Research Report Series No. 1041. 36pp.2019.
- Amaral AR, Beheregaray LB, Bilgmann K, Boutov D, Freitas L, Robertson KM, et al. Seascape genetics of a globally distributed, highly mobile marine mammal: the short-beaked common dolphin (Genus *Delphinus*). *PLoS ONE*. 2012;7(2): e31482.
- DiBattista JD, Travers MJ, Moore GI, Evans RD, Newman SJ, Feng M, et al. Seascape genomics reveals fine-scale patterns of dispersal for a reef fish along the ecologically divergent coast of Northwestern Australia. *Mol Ecol*. 2017;26(22):6206–23.
- Sandoval-Castillo J, Robinson NA, Hart AM, Strain LWS, Beheregaray LB. Seascape genomics reveals adaptive divergence in a connected and commercially important mollusc, the greenlip abalone (*Haliotis*

- laevigata*), along a longitudinal environmental gradient. *Mol Ecol.* 2018;27(7):1603–20.
37. Miller AD, Hoffmann AA, Tan MH, Young M, Ahrens C, Cocomazzo M, et al. Local and regional scale habitat heterogeneity contribute to genetic adaptation in a commercially important marine mollusc (*Halotis rubra*) from southeastern Australia. *Mol Ecol.* 2019;28(12):3053–72.
  38. Parra GJ, Bilgmann K, Peters KJ, Möller LM. Abundance and potential biological removal of common dolphins subject to fishery impacts in south Australian waters. *Front Mar Sci.* 2021;8.
  39. Goldsworthy SD, Bailleul F, NurseyBray M, Mackay A, Oxley A, Reinhold S-L, et al. Assessment of the impacts of seal populations on the seafood industry in South Australia. In: South Australian Research and Development Institute (Aquatic Sciences) A, editor. Adelaide, June 2019.
  40. Reed J, Harcourt R, New L, Bilgmann K. Extreme effects of extreme disturbances: a simulation approach to assess population specific responses. *Front Mar Sci.* 2020;7.
  41. Schumann N, Gales NJ, Harcourt RG, Arnould JPY. Impacts of climate change on Australian marine mammals. *Aust J Zool.* 2013;61(2):146.
  42. Bossart GD. Marine mammals as sentinel species for oceans and human health. *Vet Pathol.* 2011;48(3):676–90.
  43. Kemper CM, Tomo I, Bingham J, Bastianello SS, Wang J, Gibbs SE, et al. Morbillivirus-associated unusual mortality event in South Australian bottlenose dolphins is largest reported for the Southern Hemisphere. *R Soc Open Sci.* 2016;3(12): 160838.
  44. Filby NE, Bossley M, Sanderson KJ, Martinez E, Stockin KA. Distribution and population demographics of common dolphins (*Delphinus delphis*) in the Gulf St. Vincent, South Australia. *Aquat Mamm.* 2010;36(1):33–45.
  45. Costello MJ, Tsai P, Wong PS, Cheung AKL, Basher Z, Chaudhary C. Marine biogeographic realms and species endemicity. *Nat Commun.* 2017;8(1):1057.
  46. Kämpf J, Chapman P. Upwelling systems of the world: a scientific journey to the most productive marine ecosystems. Switzerland: Springer Nature; 2016. p. 443.
  47. Condie SA, Waring J, Mansbridge JV, Cahill ML. Marine connectivity patterns around the Australian continent. *Environ Model Softw.* 2005;20(9):1149–57.
  48. Ridgway KR. The 5500-km-long boundary flow off western and southern Australia. *J Geophys Res.* 2004;109(C4).
  49. Teske PR, Sandoval-Castillo J, Waters J, Beheregaray LB. An overview of Australia's temperate marine phylogeography, with new evidence from high-dispersal gastropods. *J Biogeography.* 2017;44(1):217–29.
  50. Waters JM, Wernberg T, Connell SD, Thomsen MS, Zuccarello GC, Kraft GT, et al. Australia's marine biogeography revisited: back to the future? *Austral Ecol.* 2010;35(8):988–92.
  51. York KL, Blackett MJ, Appleton BR. The Bassian Isthmus and the major ocean currents of southeast Australia influence the phylogeography and population structure of a southern Australian intertidal barnacle *Catomerus polymerus* (Darwin). *Mol Ecol.* 2008;17(8):1948–61.
  52. Eriksen RS, Davies CH, Bonham P, Coman FE, Edgar S, McEnulty FR, et al. Australia's long-term plankton observations: the integrated marine observing system national reference station network. *Front Mar Sci.* 2019;6.
  53. Dalongeville A, Benestan L, Mouillot D, Lobreaux S, Manel S. Combining six genome scan methods to detect candidate genes to salinity in the Mediterranean striped red mullet (*Mullus surmuletus*). *BMC Genomics.* 2018;19(1):217.
  54. Manel S, Schwartz MK, Luikart G, Taberlet P. Landscape genetics: combining landscape ecology and population genetics. *Trends Ecol Evol.* 2003;18(4):189–97.
  55. Oleksiak MF, Rajora OP. Population genomics: marine organisms. Switzerland: Springer Nature; 2020. p. 1–456.
  56. Robbins WD, Huveneres C, Parra GJ, Möller L, Gillanders BM. Anthropogenic threat assessment of marine-associated fauna in Spencer Gulf, South Australia. *Mar Policy.* 2017;81:392–400.
  57. Lazzarotto V. Consequences of long-term feeding trout with plant-based diets on the regulation of energy and lipid metabolism: special focus on trans-generational effects and early stages. L'Universite de Pau et des Pays de l'Adour; 2016.
  58. Sproles AE, Oakley CA, Matthews JL, Peng L, Owen JG, Grossman AR, et al. Proteomics quantifies protein expression changes in a model cnidarian colonised by a thermally tolerant but suboptimal symbiont. *ISME J.* 2019;13(9):2334–45.
  59. Tivey TR, Parkinson JE, Mandelare PE, Adressa DA, Peng W, Dong X, et al. N-linked surface glycan biosynthesis, composition, inhibition, and function in cnidarian-dinoflagellate symbiosis. *Microb Ecol.* 2020;80(1):223–36.
  60. Amarasekera M, Martino D, Ashley S, Harb H, Kesper D, Strickland D, et al. Genome-wide DNA methylation profiling identifies a folate-sensitive region of differential methylation upstream of ZFP57-imprinting regulator in humans. *FASEB J.* 2014;28(9):4068–76.
  61. Irwin RE, Thursby SJ, Ondicova M, Pentieva K, McNulty H, Richmond RC, et al. A randomized controlled trial of folic acid intervention in pregnancy highlights a putative methylation-regulated control element at ZFP57. *Clin Epigenetics.* 2019;11(1):31.
  62. Zglejc K, Franczak A. Peri-conceptual under-nutrition alters the expression of TRIM28 and ZFP57 in the endometrium and embryos during peri-implantation period in domestic pigs. *Reprod Domest Anim.* 2017;52(4):542–50.
  63. Baldwin WS, Boswell WT, Ginjupalli G, Litoff EJ. Annotation of the nuclear receptors in an Estuarine Fish species, *Fundulus heteroclitus*. *Nucl Receptor Res.* 2017;4.
  64. Jeong BC, Kang IH, Koh JT. MicroRNA-302a inhibits adipogenesis by suppressing peroxisome proliferator-activated receptor gamma expression. *FEBS Lett.* 2014;588(18):3427–34.
  65. Li L, Xie X, Qin J, Jaha GS, Saha PK, Yan J, et al. The nuclear orphan receptor COUP-TFII plays an essential role in adipogenesis, glucose homeostasis, and energy metabolism. *Cell Metab.* 2009;9(1):77–87.
  66. Allendorf FW, Hohenlohe PA, Luikart G. Genomics and the future of conservation genetics. *Nat Rev Genet.* 2010;11(10):697–709.
  67. Frankham R, Ballou JD, Briscoe DA. Introduction to conservation genetics. Cambridge University Press; 2010. p. 618.
  68. Wright S. Evolution and the genetics of populations. University of Chicago Press; 1968.
  69. Riginos C, Crandall ED, Liggins L, Bongaerts P, Trembl EA. Navigating the currents of seascape genomics: how spatial analyses can augment population genomic studies. *Curr Zool.* 2016;62(6):581–601.
  70. Kämpf J. Modelling of physical drivers of a large feeding aggregation of killer whales (*Orcinus orca*) in the western Great Australian Bight, Australia. *Deep Sea Res Part I.* 2021;171.
  71. Rajora OP. Population genomics. Concepts, approaches and applications. Springer; 2019.
  72. Wilder AP, Palumbi SR, Conover DO, Therkildsen NO. Footprints of local adaptation span hundreds of linked genes in the Atlantic silverside genome. *Evol Lett.* 2019;4(5):430–43.
  73. Crow JF, Kimura M. Introduction to population genetics theory. New York: Harper and Row; 1970. p. 591.
  74. Sunnucks P, Balkenhol N. Incorporating landscape genetics into road ecology. In: Van der Ree R, Smith D, Grilo C, editors. Handbook of road ecology. 1st ed. John Wiley and Sons; 2015.
  75. Shafer AB, Wolf JB, Alves PC, Bergstrom L, Bruford MW, Brannstrom I, et al. Genomics and the challenging translation into conservation practice. *Trends Ecol Evol.* 2015;30(2):78–87.
  76. Funk WC, Forester BR, Converse SJ, Darst C, Morey S. Improving conservation policy with genomics: a guide to integrating adaptive potential into U.S. Endangered Species Act decisions for conservation practitioners and geneticists. *Conserv Genet.* 2019;20(1):115–34.
  77. Amaral AR, Beheregaray LB, Bilgmann K, Freitas L, Robertson KM, Sequeira M, et al. Influences of past climatic changes on historical population structure and demography of a cosmopolitan marine predator, the common dolphin (genus *Delphinus*). *Mol Ecol.* 2012;21(19):4854–71.
  78. Amaral AR, Jackson JA, Möller LM, Beheregaray LB, Manuela CM. Species tree of a recent radiation: the subfamily Delphininae (Cetacea, Mammalia). *Mol Phylogenet Evol.* 2012;64(1):243–53.
  79. Amaral KBd, Amaral AR, Ewan-Fordyce R, Moreno-Benites I. Historical Biogeography of Delphininae Dolphins and Related Taxa (Artiodactyla: Delphinidae). *J Mammal Evol.* 2016;25:241.
  80. Steeman ME, Hebsgaard MB, Fordyce RE, Ho SY, Rabosky DL, Nielsen R, et al. Radiation of extant cetaceans driven by restructuring of the oceans. *Syst Biol.* 2009;58(6):573–85.

81. Manel S, Joost S, Epperson BK, Holderegger R, Storer A, Rosenberg MS, et al. Perspectives on the use of landscape genetics to detect genetic adaptive variation in the field. *Mol Ecol*. 2010;19(17):3760–72.
82. Tulloch V, Pirota V, Grech A, Crocetti S, Double M, How J, et al. Long-term trends and a risk analysis of cetacean entanglements and bycatch in fisheries gear in Australian waters. *Biodivers Conserv*. 2020;29(1):251–82.
83. Hamer DJ, Ward TM, McGarvey R. Measurement, management and mitigation of operational interactions between the South Australian Sardine Fishery and short-beaked common dolphins (*Delphinus delphis*). *Biol Conserv*. 2008;141(11):2865–78.
84. Mason S, Salgado Kent C, Bilgmann K. Common dolphins form unexpected strong social bonds: insights into social plasticity of delphinids. *Mar Mammal Sci*. 2021;37:1174.
85. Zanardo N, Bilgmann K, Parra GJ, Möller LM. Socio-genetic structure of short-beaked common dolphins in southern Australia. *J Zool*. 2016;299(2):89–97.
86. Funk WC, McKay JK, Hohenlohe PA, Allendorf FW. Harnessing genomics for delineating conservation units. *Trends Ecol Evol*. 2012;27(9):489–96.
87. Teske PR, Sandoval-Castillo J, van Sebille E, Waters J, Beheregaray LB. On-shelf larval retention limits population connectivity in a coastal broadcast spawner. *Mar Ecol Prog Ser*. 2015;532:1–12.
88. Bilgmann K, Parra GJ, Holmes L, Peters KJ, Jonsen ID, Moller LM. Abundance estimates and habitat preferences of bottlenose dolphins reveal the importance of two gulfs in South Australia. *Sci Rep*. 2019;9(1):8044.
89. Bernatchez S, Xuereb A, Laporte M, Benestan L, Steeves R, Laflamme M, et al. Seascape genomics of eastern oyster (*Crassostrea virginica*) along the Atlantic coast of Canada. *Evol Appl*. 2018;12(3):587–609.
90. Xuereb A, Benestan L, Normandeau E, Daigle RM, Curtis JMR, Bernatchez L, et al. Asymmetric oceanographic processes mediate connectivity and population genetic structure, as revealed by RADseq, in a highly dispersive marine invertebrate (*Parastichopus californicus*). *Mol Ecol*. 2018;27(10):2347–64.
91. Ward TM, McLeay LJ, Dimlich WF, Rogers PJ, McClatchie SAM, Matthews R, et al. Pelagic ecology of a northern boundary current system: effects of upwelling on the production and distribution of sardine (*Sardinops sagax*), anchovy (*Engraulis australis*) and southern bluefin tuna (*Thunnus maccoyii*) in the Great Australian Bight. *Fish Oceanogr*. 2006;15(3):191–207.
92. Ward TM, Smart J, Ivey A. Stock assessment of Australian Sardine (*Sardinops sagax*) off South Australia 2017. In: Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences) A, editor. SARDI Publication No. F2007/000765-6. SARDI Research Report Series No. 971. 107pp.2017.
93. Kool JT, Huang Z, Nichol SL. Simulated larval connectivity among Australia's southwest submarine canyons. *Mar Ecol Prog Ser*. 2015;539:77–91.
94. Short AD. Australian coastal systems: beaches, barriers and sediment compartments. Switzerland: Springer; 2020. p. 1241.
95. Ridgway KR, Godfrey JS. The source of the Leeuwin Current seasonality. *J Geophys Res: Oceans*. 2015;120(10):6843–64.
96. Bird E. Australia, Coastal Geomorphology. In: M.L. S, editor. Encyclopedia of Coastal Science Encyclopedia of Earth Science Series: Springer, Dordrecht; 2005.
97. Middleton JF, Bye JAT. A review of the shelf-slope circulation along Australia's southern shelves: cape Leeuwin to Portland. *Prog Oceanogr*. 2007;75(1):1–41.
98. Koslow JA, Pesant S, Feng M, Pearce A, Fearn P, Moore T, et al. The effect of the Leeuwin Current on phytoplankton biomass and production off Southwestern Australia. *J Geophys Res*. 2008;113(C7).
99. Currie DR, McClatchie S, Middleton JF, Nayar S. Biophysical factors affecting the distribution of demersal fish around the head of a Submarine Canyon off the Bonney Coast, South Australia. *PLoS ONE*. 2012;7(1):e30138.
100. Lourey MJ, Dunn JR, Waring J. A mixed-layer nutrient climatology of Leeuwin Current and Western Australian shelf waters: seasonal nutrient dynamics and biomass. *J Mar Syst*. 2006;59(1–2):25–51.
101. Brooke BP, Nichol SL, Huang Z, Beaman RJ. Palaeoshorelines on the Australian continental shelf: morphology, sea-level relationship and applications to environmental management and archaeology. *Cont Shelf Res*. 2017;134:26–38.
102. Kent CS, Bouchet P, Wellard R, Parnum I, Fouda L, Erbe C. Seasonal productivity drives aggregations of killer whales and other cetaceans over submarine canyons of the Bremer Sub-Basin, south-western Australia. *Australian Mammalogy*. 2020.
103. Mason S, Salgado Kent C, Donnelly D, Weir J, Bilgmann K. Atypical residency of short-beaked common dolphins (*Delphinus delphis*) to a shallow, urbanized embayment in south-eastern Australia. *R Soc Open Sci*. 2016;3(9): 160478.
104. Spitz J, Mouroucq E, Leauté J-P, Quérou J-C, Ridoux V. Prey selection by the common dolphin: fulfilling high energy requirements with high quality food. *J Exp Mar Biol Ecol*. 2010;390(2):73–7.
105. Xuereb A, Kimber CM, Curtis JMR, Bernatchez L, Fortin MJ. Putatively adaptive genetic variation in the giant California sea cucumber (*Parastichopus californicus*) as revealed by environmental association analysis of restriction-site associated DNA sequencing data. *Mol Ecol*. 2018;27(24):5035–48.
106. Manel S, Perrier C, Pralong M, Abi-Rached L, Paganini J, Pontarotti P, et al. Genomic resources and their influence on the detection of the signal of positive selection in genome scans. *Mol Ecol*. 2016;25(1):170–84.
107. Angueira AR, Shapira SN, Ishibashi J, Sampat S, Sostre-Colon J, Emmett MJ, et al. Early B cell factor activity controls developmental and adaptive thermogenic gene programming in adipocytes. *Cell Rep*. 2020;30(9):2869–78.
108. Havird JC, Mitchell RT, Henry RP, Santos SR. Salinity-induced changes in gene expression from anterior and posterior gills of *Callinectes sapidus* (Crustacea: Portunidae) with implications for crustacean ecological genomics. *Comp Biochem Physiol Part D Genomics Proteomics*. 2016;19:34–44.
109. Tabata E, Kashimura A, Kikuchi A, Masuda H, Miyahara R, Hiruma Y, et al. Chitin digestibility is dependent on feeding behaviors, which determine acidic chitinase mRNA levels in mammalian and poultry stomachs. *Sci Rep*. 2018;8(1):1461.
110. Kämpf J. Phytoplankton blooms on the western shelf of Tasmania: evidence of a highly productive ecosystem. *Ocean Sci*. 2015;11(1):1–11.
111. McClatchie S, Middleton JF, Ward TM. Water mass analysis and along-shore variation in upwelling intensity in the eastern Great Australian Bight. *J Geophys Res*. 2006;111(C8).
112. Kämpf J. On preconditioning of coastal upwelling in the eastern Great Australian Bight. *Journal of Geophysical Research*. 2010;115(C12).
113. Richardson LE, Kyser TK, James NP, Bone Y. Analysis of hydrographic and stable isotope data to determine water masses, circulation, and mixing in the eastern Great Australian Bight. *J Geophys Res*. 2009;114(C10).
114. Mackay AI, Bailleul F, Carroll EL, Andrews-Goff V, Baker CS, Bannister J, et al. Satellite derived offshore migratory movements of southern right whales (*Eubalaena australis*) from Australian and New Zealand wintering grounds. *PLoS ONE*. 2020;15(5): e0231577.
115. Möller LM, Attard CRM, Bilgmann K, Andrews-Goff V, Jonsen I, Paton D, et al. Movements and behaviour of blue whales satellite tagged in an Australian upwelling system. *Sci Rep*. 2020;10(1):21165.
116. Reeves IM, Totterdell JA, Barceló A, Sandoval-Castillo J, Batley KC, Stockin KA, et al. Population genomic structure of killer whales (*Orcinus orca*) in Australian and New Zealand waters. *Mar Mamm Sci*. 2022;38(1):151–74.
117. Cirillo A, Di Salle A, Petillo O, Melone MA, Grimaldi G, Bellotti A, et al. High grade glioblastoma is associated with aberrant expression of ZFP57, a protein involved in gene imprinting, and of CPT1A and CPT1C that regulate fatty acid metabolism. *Cancer Biol Ther*. 2014;15(6):735–41.
118. Emami NK, Jung U, Voy B, Dridi S. Radical response: effects of heat stress-induced oxidative stress on lipid metabolism in the avian liver. *Antioxidants (Basel)*. 2020;10(1):35.
119. Kämpf J, Bell D. The Murray/Coorong Estuary: meeting of the waters? In: Wolanski E, editor. *Estuaries of Australia in 2050 and beyond*. Springer, Dordrecht Heidelberg New York London: Springer; 2014. p. 31–47.
120. Bourman RP, Murray-Wallace CV, Harvey N. *Coastal Landscapes of South Australia*. Adelaide: University of Adelaide Press; 2016.
121. Waters JM. Marine biogeographical disjunction in temperate Australia: historical landbridge, contemporary currents, or both? *Divers Distrib*. 2008;14(4):692–700.

122. Colton MA, Swearer SE. Locating faunal breaks in the nearshore fish assemblage of Victoria, Australia. *Mar Freshwater Res.* 2012;63(3):218.
123. Mackay AI, Goldsworthy SD. Mitigating operational interactions with short-beaked common dolphin (*Delphinus delphis*): application of the South Australian Sardine Fishery industry Code of Practice 2016–17. In: Aquaculture RtPFa, editor. South Australian Research and Development Institute (Aquatic Sciences), Adelaide: SARDI; 2017. p. 44.
124. Breves JP, Fujimoto CK, Phipps-Costin SK, Einarsdottir IE, Björnsson BT, McCormick SD. Variation in branchial expression among insulin-like growth-factor binding proteins (igfbps) during Atlantic salmon smoltification and seawater exposure. *BMC Physiol.* 2017;17(1):2.
125. Strobel JS, Hack NL, Label KT, Cordova KL, Bersin TV, Joaurney ML, et al. Effects of food deprivation on plasma insulin-like growth factor-1 (Igf1) and Igf binding protein (Igfbp) gene transcription in juvenile cabezon (*Scorpaenichthys marmoratus*). *Gen Comp Endocrinol.* 2020;286:113319.
126. Roca-Agüjetas V, de Dios C, Leston L, Mari M, Morales A, Colell A. Recent insights into the mitochondrial role in autophagy and its regulation by oxidative stress. *Oxid Med Cell Longev.* 2019;2019:3809308.
127. Yim HS, Cho YS, Guang X, Kang SG, Jeong JY, Cha SS, et al. Minke whale genome and aquatic adaptation in cetaceans. *Nat Genet.* 2014;46(1):88–92.
128. McGowen MR, Tsagkogeorga G, Williamson J, Morin PA, Rossiter SJ. Positive selection and inactivation in the vision and hearing genes of cetaceans. *Mol Biol Evol.* 2020;37:2069.
129. Whitmarsh SK, Huveneers C, Fairweather PG. Investigating the cumulative effects of multiple stressors on fish assemblages in a semi-enclosed bay. *Marine Biol.* 2020;167(11).
130. Razgour O, Forester B, Taggart JB, Bekaert M, Juste J, Ibanez C, et al. Considering adaptive genetic variation in climate change vulnerability assessment reduces species range loss projections. *Proc Natl Acad Sci U S A.* 2019;116(21):10418–23.
131. Antao LH, Bates AE, Blowes SA, Waldock C, Supp SR, Magurran AE, et al. Temperature-related biodiversity change across temperate marine and terrestrial systems. *Nat Ecol Evol.* 2020;26:27.
132. Banks SC, Ling SD, Johnson CR, Piggott MP, Williamson JE, Beheregaray LB. Genetic structure of a recent climate change-driven range extension. *Mol Ecol.* 2010;19(10):2011–24.
133. Suthers IM, Young JW, Baird ME, Roughan M, Everett JD, Brassington GB, et al. The strengthening East Australian Current, its eddies and biological effects—an introduction and overview. *Deep Sea Res Part II Top Stud Oceanogr.* 2011;58(5):538–46.
134. Whitehead H, McGill B, Worm B. Diversity of deep-water cetaceans in relation to temperature: implications for ocean warming. *Ecol Lett.* 2008;11(11):198–207.
135. Palacios DM, Baumgartner MF, Laidre KL, Gregr EJ. Beyond correlation: integrating environmentally and behaviourally mediated processes in models of marine mammal distributions. *Endangered Species Res.* 2013;22(3):191–203.
136. Wernberg T, Russell BD, Moore PJ, Ling SD, Smale DA, Campbell A, et al. Impacts of climate change in a global hotspot for temperate marine biodiversity and ocean warming. *J Exp Mar Biol Ecol.* 2011;400(1–2):7–16.
137. Roberts CM, O'Leary BC, McCauley DJ, Cury PM, Duarte CM, Lubchenco J, et al. Marine reserves can mitigate and promote adaptation to climate change. *Proc Natl Acad Sci U S A.* 2017;114(24):6167–75.
138. Castro-Tavares D, Moura JF, Acevedo-Trejos E, Merico A. Traits shared by marine megafauna and their relationships with ecosystem functions and services. *Front Marine Sci.* 2019;6.
139. Sousa A, Alves F, Dinis A, Bentz J, Cruz MJ, Nunes JP. How vulnerable are cetaceans to climate change? Developing and testing a new index. *Ecol Indic.* 2019;98:9–18.
140. Bestley S, Ropert-Coudert Y, Bengtson Nash S, Brooks CM, Cotté C, Dewar M, et al. Marine ecosystem assessment for the southern ocean: birds and marine mammals in a changing climate. *Front Ecol Evol.* 2020;8.
141. Wild S, Krutzen M, Rankin RW, Hoppitt WJE, Gerber L, Allen SJ. Long-term decline in survival and reproduction of dolphins following a marine heatwave. *Curr Biol.* 2019;29(7):R239–40.
142. Richardson AJ, Eriksen R, Moltmann T, Hodgson-Johnston I, Wallis JR. State and Trends of Australia's Oceans. In: IMOS, editor. Hobart: Integrated Marine Observing System; 2020.
143. Foo D, Hindell M, McMahon C, Goldsworthy S, Bailleul F. Environmental drivers of oceanic foraging site fidelity in central place foragers. *Marine Biol.* 2020;167(6).
144. Watson RA, Nowara GB, Tracey SR, Fulton EA, Bulman CM, Edgar GJ, et al. Ecosystem model of Tasmanian waters explores impacts of climate-change induced changes in primary productivity. *Ecol Model.* 2013;264:115–29.
145. Batley KC, Sandoval-Castillo J, Kemper CM, Zanardo N, Tomo I, Beheregaray LB, et al. Whole genome data reveals multiple candidate genes and pathways involved in the immune response of dolphins to a highly infectious virus. *Mol Ecol.* 2021.
146. Ulman A, Zengin M, Demirel N, Pauly D. The lost fish of turkey: a recent history of disappeared species and commercial fishery extinctions for the Turkish Marmara and black seas. *Front Marine Sci.* 2020;7.
147. Daskalov GM. Overfishing drives a trophic cascade in the Black Sea. *Mar Ecol Prog Ser.* 2002;225:53–63.
148. Allendorf FW, Funk WC, Aitken SN, Byrne M, Luikart G. Conservation and the Genomics of Populations. Edition T, editor: Oxford University Press; 2022. 784 p.
149. AFMA. Small Pelagic Fishery Dolphin Strategy. Minimising dolphin interactions in the Small Pelagic Fishery. In: Australian Fisheries Management Authority (AFMA) CA, editor. Available at [afma.gov.au/sites/default/files/spf\\_dolphin\\_strategy\\_2019\\_updated\\_accessible.pdf](https://afma.gov.au/sites/default/files/spf_dolphin_strategy_2019_updated_accessible.pdf) 2019.
150. Barbosa S, Mestre F, White TA, Pauperio J, Alves PC, Searle JB. Integrative approaches to guide conservation decisions: using genomics to define conservation units and functional corridors. *Mol Ecol.* 2018;27(17):3452–65.
151. Boussarie G, Momigliano P, Robbins WD, Bonnin L, Cornu JF, Fauvelot C, et al. Identifying barriers to gene flow and hierarchical conservation units from seascape genomics: a modelling framework applied to a marine predator. *Ecography.* 2022.
152. Bilgmann K, Griffiths OJ, Allen SJ, Möller LM. A biopsy pole system for bow-riding dolphins: sampling success, behavioral responses, and test for sampling bias. *Mar Mamm Sci.* 2007;23(1):218–25.
153. Krützen M, Barre L, Möller LM, Heithaus MR, Simms C, Sherwin WB. A Biopsy System for small cetaceans: darting success and wound healing in *Tursiops* spp. *Mar Mamm Sci.* 2002;18(4):863–78.
154. Sunnucks P, Hales DF. Numerous transposed sequences of mitochondrial cytochrome oxidase I-II in aphids of the genus *Sitobion* (Hemiptera: Aphididae). *Mol Biol Evol.* 1996;13(3):510–24.
155. Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE. Double digest RAD-seq: an inexpensive method for de novo SNP discovery and genotyping in model and non-model species. *PLoS ONE.* 2012;7(5): e37135.
156. Catchen JM, Amores A, Hohenlohe P, Cresko W, Postlethwait JH. Stacks: building and genotyping Loci de novo from short-read sequences. *G3 (Bethesda).* 2011;1(3):171–82.
157. Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA. Stacks: an analysis tool set for population genomics. *Mol Ecol.* 2013;22(11):3124–40.
158. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics.* 2014;30(15):2114–20.
159. Puritz JB, Hollenbeck CM, Gold JR. dDocent: a RADseq, variant-calling pipeline designed for population genomics of non-model organisms. *PeerJ.* 2014;2: e431.
160. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. *Nat Methods.* 2012;9:357.
161. Wang J. COANCESTRY: a program for simulating, estimating and analysing relatedness and inbreeding coefficients. *Mol Ecol Resour.* 2011;11(1):141–5.
162. Attard CRM, Beheregaray LB, Moller LM. Genotyping-by-sequencing for estimating relatedness in nonmodel organisms: avoiding the trap of precise bias. *Mol Ecol Resour.* 2018;18(3):381–90.
163. Tyberghein L, Verbruggen H, Pauly K, Troupin C, Mineur F, De Clerck O. Bio-ORACLE: a global environmental dataset for marine species distribution modelling. *Glob Ecol Biogeogr.* 2012;21(2):272–81.
164. Assis J, Tyberghein L, Bosch S, Verbruggen H, Serrão EA, De Clerck O. Bio-ORACLE v2.0: extending marine data layers for bioclimatic modelling. *Global Ecol Biogeogr.* 2018;27(3):277–84.
165. Galpern P, Peres-Neto PR, Polfus J, Manseau M, Pybus O. MEMGENE: spatial pattern detection in genetic distance data. *Methods Ecol Evol.* 2014;5(10):1116–20.
166. Blanchet FG, Legendre P, Borcard D. Forward selection of explanatory variables. *Ecology.* 2008;89:2623–32.

167. Dormann CF, Elith J, Bacher S, Buchmann C, Carl G, Carré G, et al. Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. *Ecography*. 2013;36(1):27–46.
168. Prunier JG, Colyn M, Legendre X, Nimon KF, Flamand MC. Multicollinearity in spatial genetics: separating the wheat from the chaff using commonality analyses. *Mol Ecol*. 2015;24(2):263–83.
169. Prunier JG, Colyn M, Legendre X, Nimon KF, Flamand MC. Corrigendum Multicollinearity in spatial genetics: separating the wheat from the chaff using commonality analyses. *Mol Ecol*. 2017;26(21):6183.
170. O'Brien RM. A caution regarding rules of thumb for variance inflation factors. *Qual Quant*. 2007;41(5):673–90.
171. Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, et al. *Community Ecology Package*. 2.5-6 ed2019.
172. Legendre P, Oksanen J, ter Braak CJF. Testing the significance of canonical axes in redundancy analysis. *Methods Ecol Evol*. 2011;2(3):269–77.
173. Forester BR, Lasky JR, Wagner HH, Urban DL. Comparing methods for detecting multilocus adaptation with multivariate genotype–environment associations. *Mol Ecol*. 2018;27(9):2215–33.
174. Capblancq T, Forester BR. Redundancy Analysis (RDA): a Swiss Army knife for landscape genomics. *Methods Ecol Evol*. 2021;12:2298.
175. Meirmans PG, Van Tienderen PH. GENOTYPE and GENODIVE: two programs for the analysis of genetic diversity of asexual organisms. *Mol Ecol Not*. 2004;4(4):792–4.
176. Jombart T, Ahmed I. adegenet 1.3–1: new tools for the analysis of genome-wide SNP data. *Bioinformatics*. 2011;27(21):3070–1.
177. Jombart T. adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics*. 2008;24(11):1403–5.
178. Beugin MP, Gayet T, Pontier D, Devillard S, Jombart T. A fast likelihood solution to the genetic clustering problem. *Methods Ecol Evol*. 2018;9(4):1006–16.
179. Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res*. 2009;19(9):1655–64.
180. Alexander DH, Lange K. Enhancements to the ADMIXTURE algorithm for individual ancestry estimation. *BMC Bioinform*. 2011;12.
181. Weir BS, Cockerham CC. Estimating F-statistics for the analysis of population structure. *Evolution*. 1984;38(6):1358–70.
182. Benjamini Y, Yekutieli D. The control of the false discovery rate in multiple testing under dependency. *Ann Statist*. 2001;29(4):1165–88.
183. Wickham H. *ggplot2: elegant graphics for data analysis*. New York: Springer-Verlag; 2016.
184. Nam K, Lee KW, Chung O, Yim HS, Cha SS, Lee SW, et al. Analysis of the FGF gene family provides insights into aquatic adaptation in cetaceans. *Sci Rep*. 2017;7:40233.
185. NCBI. BLAST Command Line Applications User Manual. 2020.
186. Wadi L, Meyer M, Weiser J, Stein LD, Reimand J. Impact of outdated gene annotations on pathway enrichment analysis. *Nat Methods*. 2016;13(9):705–6.
187. Gene Ontology C. Gene Ontology Consortium: going forward. *Nucleic Acids Res*. 2015;43:D1049–56.
188. Cingolani P, Platts A, Le Wang L, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly*. 2012;6(2):80–92.
189. Gillespie M, Jassal B, Stephan R, Milacic M, Rothfels K, Senff-Ribeiro A, et al. The reactome pathway knowledgebase 2022. *Nucleic Acids Res*. 2022;50(D1):D687–92.

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